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ORIGINAL RESEARCH



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Influence of landscape condition on relative abundance and body condition of two generalist freshwater turtle species

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Abstract

Anthropogenic land use changes have broad impacts on biological diversity, often resulting in shifts in community composition. While many studies have documented negative impacts on occurrence and abundance of species, less attention has been given to native species that potentially benefit from anthropogenic land use changes. For many species reaching high densities in human-dominated landscapes, it is unclear whether these environments represent higher quality habitat than more natural environments. We examined the influence of landscape ecological integrity on relative abundance and body condition of two native generalist freshwater turtle species that are prevalent in anthropogenic systems, the painted turtle (Chrysemys picta) and red-eared slider (Trachemys scripta elegans). Relative abundance was negatively associated with ecological integrity for both species, but the relationship was not strongly supported for painted turtles. Body condition was positively associated with ecological integrity for painted turtles, with no strong association for red-eared sliders. Our study suggests that both species benefitted at the population level from reduced ecological integrity, but individual-level habitat quality was reduced for painted turtles. The differing responses between these two habitat generalists could partially explain why red-eared sliders have become a widespread exotic invasive species, while painted turtles have not.

KEYWORDS

Chrysemys picta, habitat generalist, land use, Trachemys scripta elegans

1 | INTRODUCTION

Anthropogenic land use changes have altered the structure and function of ecosystems on nearly all parts of the planet (Sala et al., 2000; Zwick, 1992). These alterations often reduce the ability

of landscapes to support high biological diversity and decrease the systems' resilience to environmental stressors (i.e., lower their ecological integrity; Freedman, 2015; Ordóñez & Duinker, 2012; Parrish et al., 2003). Much research has been devoted to documenting and quantifying negative impacts of anthropogenic land use changes on

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wildlife, such as declines in density and local extirpation of populations (Crawford & Bolen, 1976; Wilberg et al., 2011), lower fitness of individuals (Li et al., 2016; Slabbekoorn & Ripmeester, 2008), reduced genetic diversity (Holderegger & Di Giulio, 2010; Miraldo et al., 2016), and behavioral changes (Andersen et al., 2017; Longcore & Rich, 2004). A major outcome of broad-scale anthropogenic land use change is the loss of habitat specialists and gain of habitat generalist and synanthropic species (including invasive species), with the consequent homogenization of wildlife communities (Clavel et al., 2010; Sofaer et al., 2020).

Habitat generalists often exist at higher densities in humandominated landscapes than in more natural landscapes (Fedriani et al., 2001: Roseberry & Woolf, 1998). For example, several studies found that raccoon (Procyon lotor) densities in urban and suburban areas were higher than in adjacent rural and undisturbed areas (Prange et al., 2003; Rilev et al., 1998), Annual productivity and nesting densities of some bird species, such as Cooper's hawk (Accipiter cooperi) and American crow (Corvus brachyrhynchos), increase in urban and suburban areas (McGowan, 2001; Rosenfield et al., 1995). Anthropogenic habitat alterations can increase quality of resources such as food and cover, which provides direct benefits to some species (Bateman & Fleming, 2012; West et al., 2016). This is likely the case for strongly synanthropic species, such as raccoon (Demeny et al., 2019; Gross et al., 2012), brown rat (Rattus norvegicus; Traweger et al., 2006, Feng & Himsworth, 2014), and house sparrow (Passer domesticus; Leu et al., 2008, Khera et al., 2010). Some species can also benefit from changes in thermal conditions associated with anthropogenic land uses (Frishkoff et al., 2015; Leveau, 2018; Miles et al., 2019). For example, Bowne et al., (2018) found the proportion of females in painted turtle (Chrysemys picta) populations was positively associated with urbanization, and attributed the finding to higher soil temperatures in urban environments.

While anthropogenic land use changes can increase quality of resources for some species, many generalist species may instead benefit from reduced predation pressure (Eötvös et al., 2018; Rodewald et al., 2011), or reduced interspecific competition for resources. Competitive release occurs when the local distribution or abundance of a species increases in response to decline or extirpation of a resource competitor (Gause, 1932; Hardin, 1960). Many experimental and observational studies have confirmed potential for competitive release (Berger & Gese, 2007; Hairston, 1986; Menge, 1976; Segre et al., 2016). In the context of reduced predation or competition pressure, anthropogenic habitat alterations could both reduce quality of resources and result in increased densities of generalist species (Cruz-Elizalde et al., 2016; Decena et al., 2020; Peltzer et al., 2006).

In the absence of pre- and post-anthropogenic habitat alteration community data, health of individuals can provide insights into whether generalist species benefit from anthropogenic habitat alterations. Specifically, body condition index (BCI) can be a useful metric to assess habitat quality (Maceda-Veiga et al., 2014; Pulliam, 2000; Sasaki et al., 2016). A BCI score represents the relationship between the weight and size of an individual relative to the study group, typically using residuals from a log-transformed length—weight regression

(Schulte-Hostedde et al., 2005). Individuals with BCI scores above the mean have above average amounts of metabolizable tissue (fat or protein) relative to their length and vice versa (Schulte-Hostedde et al., 2005). Body condition correlates with fitness metrics such as survival probability and fecundity (Bender et al., 2008; Burton et al., 2006; Carranza & Hidalgo de Trucios, 1993).

The painted turtle (Chrysemys picta) and red-eared slider (Trachemys scripta elegans) are generalist freshwater turtle species native to North America (Ernst & Lovich, 2009). Our focal subspecies, eastern painted turtle [C. p. picta] and midland painted turtle [C. p. marginata], are widely distributed across much of the eastern United States and southeastern Canada (Ernst & Lovich, 2009). The red-eared slider, a subspecies of the pond slider (*T. scripta*), is native to a large portion of the east-central United States (Ernst & Lovich, 2009). However, due to their popularity in the pet trade and ability to persist in a wide variety of environmental conditions, nonnative populations of red-eared sliders have become established in many regions of the world (Héritier et al., 2017; Lambert et al., 2019), and it is considered one of the world's worst invasive species (Lowe et al., 2000). Both species generally prefer shallow lentic freshwater habitats containing a soft mucky bottom with abundant aquatic plants (DonnerWright et al., 1999; Janzen et al., 1992; Morreale & Gibbons, 1986). Both species are also commonly found in wetlands associated with anthropogenic land use, such as agricultural farm ponds and urban retention ponds (Buchanan et al., 2019; Stone et al., 2005). Further, many studies have indicated that densities of painted turtles and red-eared sliders in anthropogenic wetlands are much higher than other turtle species occupying the same wetlands (Brown, Farallo, et al., 2011; Failey et al., 2007; Glorioso et al., 2010).

Although generalist turtle species can achieve high densities in human-dominated landscapes, little research has been conducted to assess whether these environments represent higher quality habitat than more natural systems. The purpose of this study was to determine whether relative abundance and body condition of painted turtles sampled in West Virginia, and red-eared sliders sampled in Texas, are correlated with ecological integrity of the surrounding landscape. We hypothesized that relative abundance of these species would be negatively correlated with ecological integrity, which would suggest that human-dominated landscapes can support larger populations, potentially due to reduced predation or competition pressure. We also hypothesized that BCI score for these species would be negatively correlated with ecological integrity, which would suggest that habitat quality for these species is better in human-dominated landscapes.

2 | METHODS

2.1 | Species data and sampling sites

We collated turtle capture and measurement data previously collected by the authors for painted turtles in West Virginia and redeared sliders in Texas. The data were originally collected for a wide variety of research projects primarily focused on relationships between relative abundance and land use and management (Brown et al., 2012; Gulette, 2018; Mali et al., 2013; Watson & Pauley, 2006) and investigations of hoop net sampling methodology (Gulette et al., 2019; Mali et al., 2014; Oxenrider et al., 2019). For all study sites, turtle populations were sampled using hoop net traps, primarily baited with canned sardines. Turtles were sampled throughout the active season (March-September) in both states. Trap size varied based on study objectives and ranged from 0.3 to 0.91 m diameter in hoop width.

Trapping occurred between 1999 and 2019 in West Virginia, and between 2008 and 2013 in Texas (Appendix S1). Painted turtles were sampled at 49 wetlands across 10 counties in southern and eastern West Virginia (Figure 1; Appendix S1). Red-eared sliders were sampled at 43 wetlands across five counties in south, central, and west Texas (Figure 1; Appendix S1). Midline carapace length (MCL) was measured to the nearest 1 mm using tree calipers (method D in

Iverson & Lewis, 2018). Weight was measured using spring scales to the nearest 1, 5, 10, and 50 g for turtles weighing \leq 10, \leq 600, \leq 2,500, and >2,500 g, respectively (Brown et al., 2020). Turtles were individually marked using marginal scute notches (Cagle, 1939). In both states, sampled wetlands occurred in agricultural systems, river backwaters, and natural areas. In Texas, several wetlands also occurred in heavily urban environments. Wetlands ranged in size from 0.008 to 5.577 ha (median = 0.063 ha) in West Virginia and 0.018 to 66.264 ha (median = 1.145 ha) in Texas.

2.2 | Landscape condition

We used NatureServe's Landscape Condition Model (LCM) for Temperate North America as our landscape condition index (Hak & Comer. 2017). This index is based on 20 landscape characteristics, categorized as transportation (including roads at multiple

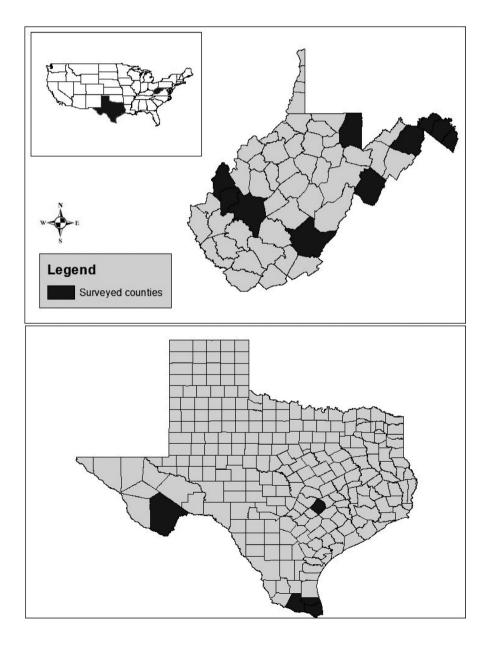


FIGURE 1 Map of counties sampled for painted turtles (Chrysemys picta) in West Virginia (top) and red-eared sliders (Trachemys scripta elegans) in Texas (bottom), USA (inset). We sampled 49 wetlands in West Virginia between 1999 and 2019, and 43 wetlands in Texas between 2008 and 2013

classification levels), urban and industrial development, and managed and modified land cover (Hak & Comer, 2017). Ecological integrity is scored from 0 to 1, with scores close to 0 and 1 representing areas of lowest and highest ecological integrity, respectively (Hak & Comer, 2017). The model has a spatial resolution of 90 m². We digitized sampled wetland boundaries using aerial imagery and created buffers around each water body at 0.1 km, 1.0 km, and 2.5 km. The 0.1, 1.0, and 2.5 km buffers were specified to approximate local wetland, home range size, and dispersal area size buffers for our focal turtle species (Bodie & Semlitsch, 2000; Bowne & White, 2004; Gibbons, 1967; Tucker & Lamer, 2008). We computed the mean landscape condition value within each buffer.

The LCM provides a static measure of ecological integrity and was constructed using datasets representing environmental conditions between approximately 2003 and 2010, including the 2011 National Landcover Database (NLCD; Hak & Comer, 2017). We performed a preliminary analysis to ensure the land cover surrounding our sites was similar between the turtle sampling and LCM timeframes. We obtained the NLCD data for the years 2001, 2006, 2011, and 2016, clipped the layers to a 2.5 km buffer around each site, and computed Pearson's correlation coefficients between the 2011 NLCD layer and the layer that most closely matched the turtle sampling timeframe. Pearson's correlation coefficients were ≥0.9588 across all sites, indicating the LCM likely provides a reliable metric of ecological integrity for our sites. All spatial analyses were performed using ArcMAP 10.6 (ESRI, Redlands, California, USA).

2.3 | Turtle relative abundance

Trapping effort varied among wetlands, and thus, we used captures-per-unit-effort (CPUE) as our metric of relative abundance (Brown et al., 2011; Murray & Seed, 2010). To obtain CPUE, we divided the number of unique individual captures of the focal species by the number of trap days (TD) at each wetland, where a single TD represented one trap in the water for one day (Appendix S1). To minimize potential CPUE biases due to level of trapping effort, we specified a target effort of 50 TD and removed sites with <20 TD (n = 2). For sites with >50 TD, we excluded all subsequent days of trapping once the site reached 50 TD. For the analysis, site-level TD ranged from 20 to 123 (mean = 54, hereafter reduced CPUE analysis; Appendix S1). We supplemented the reduced CPUE analysis with an additional analysis that included all TD at each site (hereafter full CPUE analysis; Appendices S2 and S3). For both analyses, we excluded 9 sites in West Virginia because number of trap days was not available.

2.4 | Turtle body condition

We computed turtle BCI scores using the residuals of log-transformed MCL-weight regressions (Schulte-Hostedde et al., 2005). We computed BCIs separately for each sex within each species and standardized the values (0 mean, 1 standard deviation) so that BCI scores were

weighted equally among each sex and species (Schulte-Hostedde et al., 2005). We removed adult and subadult turtle capture records with unrecorded sex from the dataset. We also excluded juveniles, including painted turtles <89 mm MCL (Balcombe & Licht, 1987; Lefevre & Brooks, 1995) and red-eared sliders <101 mm MCL (Cagle, 1948) because sex was unknown for this size class.

2.5 | Statistical analyses

We used linear mixed-effects models to analyze the relationship between landscape condition and turtle CPUE and BCI (Zuur et al., 2009). For CPUE, we grouped wetlands into four size classes (Class 1 = <2.750 ha; Class 2 = 2.750 ha < 10.795 ha; Class 3 = 10.795 ha < 33.615 ha; Class $4 = \ge 35.615 \text{ ha}$) using the Jenks natural breaks classification method (Jenks, 1977). This method is based on Fisher's "Exact Optimization" method (Fischer, 1958), which seeks to optimize homogeneity within groups by minimizing the sum of squares difference. We included size class as a random effect to account for potential effects of wetland size on CPUE. Fixed effects included landscape condition value (LCV) and trap size. For this analysis, we used the LCV extent that was most supported for the BCI analysis. We specified traps as small (0.3 m) or large (0.76-0.91 m) to account for potential trap size effects on CPUE of painted turtles in West Virginia. Trap size (0.76 m) was consistent for all wetlands sampled for red-eared sliders in Texas. For CPUE, preliminary analyses indicated the LCV relationship may be quadratic, and thus we tested LCV as both a linear and quadratic predictor. For BCI, we included wetland as a random effect to account for site-level environmental variation independent of landscape condition that could influence BCI. Fixed effects included sex and mean LCV surrounding the wetland at distances of 0.1, 1.0, and 2.5 km. We tested the influence of sex as both an additive effect and an interactive effect.

We used Akaike's information criterion corrected for small sample size (AIC_c) to rank candidate models (Burnham & Anderson, 2004). We considered models to have strong support if $\Delta AICc < 2$ (Burnham et al., 2011). For the most supported models, we assessed confidence for an effect of each variable by computing the 85% confidence intervals (CI) of the beta coefficients (Arnold, 2010) and considered there to be evidence for a strong effect when CIs did not overlap zero (Halsey, 2019). For all analyses, we assessed assumptions of normality using quantile-quantile plots and homoscedasticity using residual plots (Zuur et al., 2009, 2010). For the CPUE models, we removed one painted turtle site to satisfy the assumption of normality. For the BCI models, we removed 23 extreme outliers (>4 standard deviations from the mean), which likely represented incorrect MCL or weight measurements. All analyses were conducted using program R (version 3.6.3). We performed the Jenks natural breaks classification using the package BAMMtools (version 2.1.7) and assessed model assumptions using the package car (version 3.0-6). We created mixed-effects models using the package nlme (version 3.1-142), performed model selection analyses using the package AICcmodavg (version 2.2-2), and plotted results using the package ggplot2 (version 3.2.1).

RESULTS

For the reduced CPUE dataset, CPUE per wetland ranged from 0.02 to 1.00 (mean = 0.22) for painted turtles in West Virginia and from 0 to 0.70 (mean = 0.14) for red-eared sliders in Texas (Appendix S1). For the BCI dataset, unique turtle captures per wetland ranged from 1 to 109 (mean = 14) for painted turtles in West Virginia and from 1 to 135 (mean = 17) for red-eared sliders in Texas (Appendix S1). The LCV scores ranged from 0.016 to 0.6 (mean = 0.221) for sampled wetlands in West Virginia and from 0.005 to 0.890 (mean = 0.437) for sampled wetlands in Texas. The LCV scores were highly correlated among the three buffer sizes within each state ($r^2 = .82-.96$), indicating landscape condition near the wetland was similar to landscape condition in the surrounding landscape, at least at the spatial resolution of the LCM.

For the painted turtle reduced CPUE analysis, the most supported model was the null model ($w_i = 0.51$; Table 1). The second most supported model was the linear 2.5 km LCV model ($w_i = 0.26$, Δ AIC_c = 1.31). For this model, predicted CPUE decreased by 0.296 as LCV increased from 0 to 1 (Figure 2a), but the CI broadly overlapped zero (-0.695-0.104). We obtained similar results for the full CPUE analysis (Appendices S2 and S3). For painted turtle BCI, the linear 2.5 km LCV model was the most supported model ($w_i = 0.28$; Table 1). The linear 2.5 km LCV + sex ($w_i = 0.18$, $\Delta AIC_c = 0.90$) and linear 1.0 km LCV ($w_i = 0.13$, $\Delta AIC_c = 1.51$) models also had strong support. The null model received the lowest support ($w_i = 0.01$; Table 1). For the most supported model, predicted BCI increased 1.32 standard deviations as LCV increases from 0-1 (Figure 3a), and the CI did not overlap zero (0.782-1.849).

For the red-eared slider reduced CPUE analysis, the most supported model was the linear 2.5 km LCV model ($w_i = 0.53$), but the quadratic 2.5 km LCV ($w_i = 0.25$, $\Delta AIC_c = 1.49$) and null ($w_i = 0.22$, $\Delta AIC_c = 1.76$) models also had strong support (Table 1). The linear 2.5 km LCV model-predicted CPUE decreased 0.19 standard deviations as LCV increased from 0 to 1, and 85% CI did not overlap zero (-0.058 to -0.328; Figure 2b). The modeled relationship was similar for the full CPUE analysis (Appendix S3), except the quadratic 2.5 km LCV model received higher support than the linear model (Appendix S2). For red-eared slider BCI, the most supported model was the null model ($w_i = 0.36$; Table 1). The linear 2.5 km LCV ($w_i = 0.16$, $\Delta AIC_c = 1.61$) and linear 1.0 km LCV ($w_i = 0.13$, $\Delta AIC_c = 1.99$) models also had strong support. For the linear 2.5 km LCV model, predicted BCI increased 0.16 standard deviations as LCV increases from 0 to 1 (Figure 3b). However, the 85% CI broadly overlapped zero (-0.174 - 0.503).

DISCUSSION

Generalist species often appear to benefit from anthropogenically degraded landscapes, but the underlying causes that enable increased densities of most habitat generalists are not clear. We sought to improve our understanding of the underlying forces

TABLE 1 Model selection results for the influence of landscape integrity (landscape condition value [LCV]) on captures-per-uniteffort (CPUE) and body condition index (BCI) of painted turtles (Chrysemys picta) in West Virginia and red-eared sliders (Trachemys scripta elegans) in Texas

Model	AIC _c	ΔAIC_c	14/
	AIC	AAIC	w _i
Chrysemys picta CPUE			
(.)	2.08	0.00	0.51
LCV _{2.5}	3.39	1.31	0.26
	5.08	3.00	0.20
$LCV_{2.5}$ (Q) $LCV_{2.5}$ + Trap size	5.82	3.74	0.08
$LCV_{2.5} + Trap size$ LCV _{2.5} (Q) + Trap size	7.56	5.48	0.03
BCI	7.50	5.40	0.00
LCV _{2.5}	1,707.23	0.00	0.28
LCV _{2.5} + Sex	1,708.13	0.90	0.18
LCV _{1.0}	1,708.74	1.51	0.13
LCV _{0.1}	1,708.78	1.56	0.13
$LCV_{2.5} \times Sex$	1,709.46	2.23	0.09
LCV _{1.0} + Sex	1,709.99	2.76	0.07
LCV _{0.1} + Sex	1,710.21	2.98	0.06
LCV _{1.0} × Sex	1,711.51	4.28	0.03
LCV _{0.1} × Sex	1,712.12	4.89	0.02
(.)	1,714.15	6.93	0.01
Trachemys scripta elega	ns		
CPUE			
LCV _{2.5}	-31.25	0.00	0.53
LCV _{2.5} (Q)	-29.75	1.49	0.25
(.)	-29.48	1.76	0.22
BCI			
(.)	2,011.47	0.00	0.36
LCV _{2.5}	2,013.08	1.61	0.16
LCV _{1.0}	2,013.47	1.99	0.13
LCV _{0.1}	2,013.49	2.02	0.13
$LCV_{2.5} + Sex$	2,015.10	3.63	0.06
$LCV_{1.0} + Sex$	2,015.49	4.01	0.05
$LCV_{0.1} + Sex$	2,015.52	4.04	0.05
$LCV_{2.5} \times Sex$	2,017.13	5.66	0.02
$LCV_{1.0} \times Sex$	2,017.49	6.02	0.02
$LCV_{0.1} \! imes \! Sex$	2,017.54	6.07	0.02

Note: For CPUE, we used a reduced trapping dataset with a target of 50 trap days per site. We used Akaike's information criterion corrected for small sample size (AIC_c) to rank candidate models. For CPUE, we used the 2.5 km LCV and tested a linear and quadratic (Q) relationship. The size of traps (Trap Size) varied at West Virginia sites and was included as a candidate predictor for C. picta. For BCI, we ranked mean LCV at 0.1, 1.0, and 2.5 km surrounding wetlands. We also tested the influence of sex as an additive and interactive predictor at the three spatial scales. We standardized BCI by species and sex prior to analysis. The null model is shown as (.) and includes only the intercept. Wetland buffer distance is denoted by subscripts following the LCV term. Akaike weights are represented as w_i.

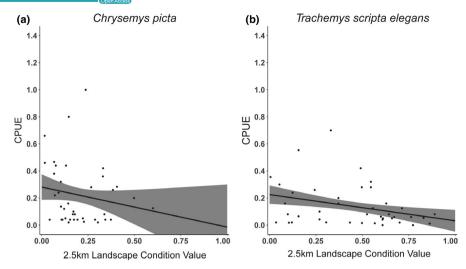


FIGURE 2 Model-estimated relationship between mean 2.5 km landscape condition value (LCV) and captures-per-unit-effort (CPUE) for (a) 39 painted turtle (*Chrysemys picta*) wetlands located across 10 counties in West Virginia, and (b) 41 red-eared slider (*Trachemys scripta elegans*) wetlands located across five counties in Texas using the reduced CPUE analysis dataset. Wetlands where trap days could not be calculated were excluded from this analysis. We included wetland size as a random effect in analyses to account for the influence of size on CPUE. Black circles depict observed CPUE, and gray bands depict 85% confidence intervals. Note the maximum LCV for *C. picta* sites was 0.6

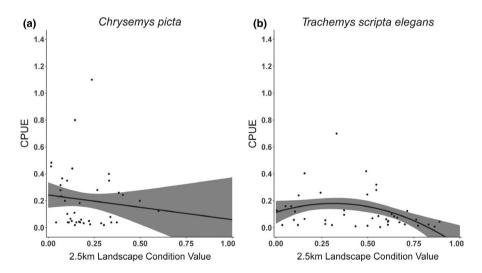


FIGURE 3 Model-estimated relationships between mean 2.5 km landscape condition value (LCV) and standardized body condition index (BCI) scores for (a) painted turtles (*Chrysemys picta*; n = 625) sampled at 46 wetlands across 10 counties in West Virginia and (b) red-eared sliders (*Trachemys scripta elegans*; n = 715) sampled at 42 wetlands across 5 counties in Texas. We included wetland as a random effect in analyses to account for site-level environmental variation independent of landscape condition that could influence BCI. Black circles depict standardized BCI values, and gray bands depict 85% confidence intervals. Note the maximum LCV for *C. picta* sites was 0.6

allowing two generalist turtle species to maintain high abundances in highly degraded systems. Our results provide some support for the hypothesis that generalist freshwater turtle species benefit from anthropogenic land use, as relative abundance of both species was negatively associated with ecological integrity. However, the two species differed in individual-level responses, with reduced ecological integrity appearing to negatively impact painted turtles but not red-eared sliders. This indicates that painted turtles may benefit from anthropogenic land uses through other factors than improved habitat quality, such as reduced predation or competition pressure, which has been documented for other freshwater turtle species (Petrozzi et al., 2021; Ryan et al., 2008; Spencer & Thompson, 2005).

It is interesting that individual-level responses to ecological integrity differed between these two generalist turtle species, as both species are prevalent in wetlands that span a wide range of environmental conditions (Brown et al., 2012; Buchanan et al., 2019). Wetlands associated with anthropogenic landscapes generally differ from those in more natural systems. For example, wetlands associated with developed and working lands are often more eutrophic (Kennish, 2002; Smith & Schindler, 2009), which in turn influences many abiotic and biotic factors (McCormick & Laing, 2003; McGoff et al., 2013; Naselli-Flores & Barone, 2000). Created wetlands (e.g., farm ponds and mitigation wetlands) also tend to be deeper than natural wetlands (Cole & Brooks, 2000; Cole et al., 2006; Gamble &

Mitsch, 2009). These wetland-specific characteristics likely interact with the surrounding landscape condition to influence habitat quality for the two species (Buchanan et al., 2019; Cosentino et al., 2010; Ryan et al., 2008). While our study was not designed to control for wetland characteristics, we encourage future studies to explore interactions between landscape integrity and species-specific wetland habitat quality.

The red-eared slider has successfully established non-native populations in many regions of the world (e.g., France [García-Díaz et al., 2017], Japan [Kakuda et al., 2019], South Korea [Oh et al., 2017]). Our results suggest that even within their native distribution (apart from one study site in west Texas), red-eared sliders benefit from environmental conditions associated with lower ecological integrity. The ability to exploit anthropogenic habitats, in conjunction with potentially reduced competitive pressure in anthropogenically altered systems (Cadi & Joly, 2003), could explain why red-eared sliders are a particularly successful invasive species. In contrast, painted turtles did not appear to strongly benefit from lower ecological integrity and are also not a prominent invasive species, despite also being common in the pet trade (Hohn, 2003; Telecky, 2001). Interestingly, red-eared sliders typically achieve much higher densities than painted turtles in sympatric areas (Bodie et al., 2000; Dreslik et al., 2005), indicating red-eared sliders may be competitively dominant (Lindeman, 1999; Polo-Cavia et al., 2011).

Anthropogenic land use changes result in creation, loss, and alteration of environmental conditions, resulting in wildlife species "winners and losers" (McKinney & Lockwood, 1999). Globally, freshwater turtles are declining in human-dominated systems due to a variety of pressures, such as habitat loss and degradation, and overexploitation for food or pets (Gibbons et al., 2000; Lovich et al., 2018). Further, the general life history strategy of freshwater turtles is characterized by a long lifespan, delayed sexual maturity, and low annual recruitment (Congdon et al., 1994), which can result in both slow declines and slow recovery rates (e.g., Howell et al., 2019; Mullin et al., 2020). Our investigation of the relationship between landscape integrity and habitat quality for two widely distributed habitat generalist turtles in North America suggests that ecological integrity has little influence on habitat quality for the red-eared slider, potentially explaining its prominence as an exotic invasive species (Lowe et al., 2000), and ecological degradation could benefit both species at the population level. Thus, as many regions in North America continue to shift toward heavy anthropogenic use (e.g., agriculture and urbanization; Brown et al., 2005; Ordonez et al., 2014), we expect these two species to be "winners" in comparison with other sympatric freshwater turtle species.

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CONFLICT OF INTEREST

The authors declare no competing interests or conflicts of interest.

AUTHOR CONTRIBUTION

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DATA AVAILABILITY STATEMENT

The data used in this study are achieved in the Dryad data repository: https://doi.org/10.5061/dryad.jdfn2z39x.

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REFERENCES

- Andersen, G. E., Johnson, C. N., Barmuta, L. A., & Jones, M. E. (2017). Use of anthropogenic linear features by two medium-sized carnivores in reserved and agricultural landscapes. *Scientific Reports*, 7, 1–12. https://doi.org/10.1038/s41598-017-11454-z
- Arnold, T. W. (2010). Uninformative parameters and model selection using Akaike's Information Criterion. *Journal of Wildlife Management*, 74, 1175–1178. https://doi.org/10.1111/j.1937-2817.2010.tb012 36.x
- Balcombe, J. P., & Licht, L. E. (1987). Some aspects of the ecology of the midland painted turtle, *Chrysemys picta marginata* in Wye Marsh, Ontario. *Canadian Field-Naturalist*, 101, 98–100.
- Bateman, P. W., & Fleming, P. A. (2012). Big city life: Carnivores in urban environments. *Journal of Zoology*, 287, 1–23. https://doi.org/10.1111/j.1469-7998.2011.00887.x
- Bender, L. C., Cook, J. G., Cook, R. C., & Hall, P. B. (2008). Relations between nutritional condition and survival of North American elk *Cervus elaphus. Wildlife Biology*, 14, 70–80.
- Berger, K. M., & Gese, E. M. (2007). Does interference competition with wolves limit the distribution and abundance of coyotes? *Journal of Animal Ecology*, 76, 1075–1085. https://doi.org/10.1111/j.1365-2656.2007.01287.x
- Bodie, J. R., & Semlitsch, R. D. (2000). Spatial and temporal use of floodplain habitats by lentic and lotic species of aquatic turtles. *Oecologia*, 122, 138–146. https://doi.org/10.1007/PL00008830
- Bodie, J. R., Semlitsch, R. D., & Renken, R. B. (2000). Diversity and structure of turtle assemblages: Associations with wetland characters across a floodplain landscape. *Ecography*, 23, 444–456. https://doi.org/10.1111/j.1600-0587.2000.tb00301.x
- Bowne, D. R., Cosentino, B. R., Anderson, L. J., Blotch, C. P., Cooke, S., Crumrine, P. W., Dallas, J., Doran, A., Dosch, J. H., Druckenbrod, D. L., Durtche, R. D., Garneau, D., Genet, K. S., Fredericksen, T. S., Kish, P. A., Kolozsvary, M. B., Kuserk, F. T., Lindquist, E. S., Mankiewicz, C., ... Zimmermann, C. R. (2018). Effects of urbanization on the population structure of freshwater turtles across the United States. Conservation Biology, 32, 1150-1161. https://doi.org/10.1111/cobi.13136
- Bowne, D. R., & White, H. R. (2004). Searching strategy of the painted turtle *Chrysemys picta* across spatial scales. *Animal Behaviour*, *68*, 1401–1409. https://doi.org/10.1016/j.anbehav.2004.01.018
- Brown, D. J., Farallo, V. R., Dixon, J. R., Baccus, J. T., Simpson, T. R., & Forstner, M. R. J. (2011). Freshwater turtle conservation in Texas: Harvest effects and efficacy of the current management regime. *Journal of Wildlife Management*, 75, 486-494. https://doi.org/10.1002/jwmg.73
- Brown, D. G., Johnson, K. M., Loveland, T. R., & Theobald, D. M. (2005). Rural land-use trends in the conterminous United States, 1950-2000. *Ecological Applications*, 15, 1851-1863. https://doi.org/10.1890/03-5220
- Brown, D. J., Mali, I., & Forstner, M. R. J. (2011). No difference in short-term temporal distribution of trapping effort on hoop-net capture efficiency for freshwater turtles. *Southeastern Naturalist*, 10, 245–250. https://doi.org/10.1656/058.010.0205
- Brown, D. J., Mali, I., Jones, M. C., & Forstner, M. R. J. (2020). Morphometric data for five freshwater turtles in south, central, and west Texas. *Data in Brief*, 29, 105356.
- Brown, D. J., Schultz, A. D., Dixon, J. R., Dickerson, B. E., & Forstner, M. R. J. (2012). Decline of red-eared sliders (*Trachemys scripta elegans*) and Texas spiny softshells (*Apalone spinifera emoryi*) in the Lower Rio

- Grande Valley of Texas. *Chelonian Conservation and Biology*, 11, 138–143. https://doi.org/10.2744/CCB-0928.1
- Buchanan, S. W., Buffum, B., Puggioni, G., & Karraker, N. E. (2019). Occupancy of freshwater turtles across a gradient of altered land-scapes. *Journal of Wildlife Management*, 83, 435-445. https://doi.org/10.1002/jwmg.21596
- Burnham, K. P., & Anderson, D. R. (2004). Multimodel inference: Understanding AIC and BIC model selection. Sociological Methods and Research, 33, 261–304. https://doi.org/10.1177/0049124104 268644
- Burnham, K. P., Anderson, D. R., & Huyvaert, K. P. (2011). AIC model selection and multimodel inference in behavioral ecology: Some background, observations, and comparisons. *Behavioral Ecology* and Sociobiology, 65, 23–35. https://doi.org/10.1007/s0026 5-010-1029-6
- Burton, N. H. K., Rehfisch, M. M., Clark, N. A., & Dodd, S. G. (2006). Impacts of sudden winter habitat loss on the body condition and survival of redshank *Tringa totanus*. *Journal of Applied Ecology*, 43, 464–473. https://doi.org/10.1111/j.1365-2664.2006.01156.x
- Cadi, A., & Joly, P. (2003). Competition for basking places between the endangered European pond turtle (*Emys orbicularis galloitalica*) and the introduced red-eared slider (*Trachemys scripta elegans*). Canadian Journal of Zoology, 81, 1392–1398.
- Cagle, F. R. (1939). A system of marking turtles for future identification. *Copeia*, 1939, 170–173. https://doi.org/10.2307/1436818
- Cagle, F. R. (1948). Sexual Maturity in the male turtle, *Pseudemys scripta* troostii. Copeia, 1948, 108–111. https://doi.org/10.2307/1438413
- Carranza, J., & Hidalgo de Trucios, S. J. (1993). Condition-dependence and sex traits in the male great bustard. *Ethology*, 94, 187–200. https://doi.org/10.1111/j.1439-0310.1993.tb00559.x
- Clavel, J., Julliard, R., & Devictor, V. (2010). Worldwide decline of specialist species: Toward a global functional homogenization? Frontiers in Ecology and the Environment, 9, 222–228. https://doi. org/10.1890/080216
- Cole, C. A., & Brooks, R. P. (2000). A comparison of the hydrologic characteristics of natural and created mainstem floodplain wetlands in Pennsylvania. *Ecological Engineering*, 14, 221–231. https://doi.org/10.1016/S0925-8574(99)00004-X
- Cole, C. A., Urban, C. A., Russo, P., Murray, J., Hoyt, D., & Brooks, R. P. (2006). Comparison of the long-term water levels of created and natural reference wetlands in northern New York, USA. *Ecological Engineering*, 27, 166–172. https://doi.org/10.1016/j.ecoleng.2006.03.003
- Congdon, J. D., Dunham, A. E., & Van Loben Sels, R. C. (1994). Demographics of common snapping turtles (*Chelydra serpentina*): Implications for conservation and management of long-lived organisms. *American Zoologist*, 34, 397–408.
- Cosentino, B. J., Schooley, R. L., & Phillips, C. A. (2010). Wetland hydrology, area, and isolation influence occupancy and spatial turnover of the painted turtle, *Chrysemys picta*. *Landscape Ecology*, *25*, 1589–1600. https://doi.org/10.1007/s10980-010-9529-3
- Crawford, J. A., & Bolen, E. G. (1976). Effects of land use on lesser prairie chickens in Texas. *Journal of Wildlife Management*, 40, 96–104. https://doi.org/10.2307/3800160
- Cruz-Elizalde, R., Berriozabal-Islas, C., Hernández Salinas, U., Martínez-Morales, M. A., & Ramírez-Bautista, A. (2016). Amphibian species richness and diversity in a modified tropical environment of central Mexico. *Tropical Ecology*, *57*, 407–417.
- Decena, S. Y. P., Avorque, C. A., Decena, I. C. P., Asis, P. D., & Pacle, B. (2020). Impact of habitat alteration on amphibian diversity and species composition in a lowland tropical rainforest in Northeastern Leyte, Philippines. Scientific Reports, 10, 10547.
- Demeny, K., McLoon, M., Winesett, B., Fastner, J., Hammerer, E., & Pauli, J. N. (2019). Food subsidies of raccoons (*Procyon lotor*) in anthropogenic landscapes. *Canadian Journal of Zoology*, *97*, 654–657.

- DonnerWright, D. M., Bozek, M. A., Probst, J. R., & Anderson, E. M. (1999). Responses of turtle assemblage to environmental gradients in the St. Croix River in Minnesota and Wisconsin, U.S.A. Canadian Journal of Zoology, 77, 989–1000.
- Dreslik, M. J., Kuhns, A. R., & Phillips, C. A. (2005). Structure and composition of a southern Illinois freshwater turtle assemblage. *Northeastern Naturalist*, 12, 173–186.
- Eötvös, C. B., Magura, T., & Lövei, G. L. (2018). A meta-analysis indicates reduced predation pressure with increasing urbanization. *Landscape* and Urban Planning, 180, 54–59. https://doi.org/10.1016/j.landu rbplan.2018.08.010
- Ernst, C. H., & Lovich, J. E. (2009). Turtles of the United States and Canada (2nd ed.). Johns Hopkins University Press.
- Failey, E. L., McCoy, J. C., Price, S. J., & Dorcas, M. E. (2007). Ecology of turtles inhabiting golf course and farm ponds in the western Piedmont of North Carolina. *Journal of the North Carolina Academy of Science*, 123, 221–232.
- Fedriani, J. M., Fuller, T. K., & Sauvajot, R. M. (2001). Does availability of anthropogenic food enhance densities of omnivorous mammals? An example with coyotes in southern California. *Ecography*, 24, 325–331. https://doi.org/10.1034/j.1600-0587.2001.240310.x
- Feng, A. Y. T., & Himsworth, C. G. (2014). The secret life of the city rat: A review of the ecology of urban Norway and black rats (*Rattus norvegicus* and *Rattus rattus*). *Urban Ecosystems*, 17, 149–162. https://doi.org/10.1007/s11252-013-0305-4
- Fischer, W. D. (1958). On grouping for maximum homogeneity. *Journal of the American Statistical Association*, *53*, 789–798. https://doi.org/10.1080/01621459.1958.10501479
- Freedman, B. (2015). Ecological effects of environmental stressors. In Oxford research Encyclopedia of environmental science (pp. 1–29).
- Frishkoff, L. O., Hadly, E. A., & Daily, G. C. (2015). Thermal niche predicts tolerance to habitat conversion in tropical amphibians and reptiles. *Global Change Biology*, *31*, 3901–3916. https://doi.org/10.1111/gcb.13016
- Gamble, D. L., & Mitsch, W. J. (2009). Hydroperiods of created and natural vernal pools in central Ohio: A comparison of depth and duration of inundation. *Wetlands Ecology and Management*, *17*, 385–395. https://doi.org/10.1007/s11273-008-9115-5
- García-Díaz, P. G., Ramsey, D. S. L., Woolnough, A. P., Franch, M., Llorente, G. A., Montori, A., Buenetxea, X., Larrinaga, A. R., Lasceve, M., Álvarez, A., Traverso, J. M., Valdeón, A., Crespo, A., Rada, V., Ayllón, E., Sancho, V., Lacomba, J. I., Bataller, J. V., & Lizana, M. (2017). Challenges in confirming eradication success of invasive red-eared sliders. *Biological Invasions*, 19, 2739–2750. https://doi.org/10.1007/s10530-017-1480-7
- Gause, G. F. (1932). Experimental studies on the struggle for existence: Mixed population of two species of yeast. *Journal of Experimental Biology*, 9, 389–402.
- Gibbons, J. W. (1967). Reproductive potential, activity, and cycles in the painted turtle, *Chrysemys picta*. *Ecology*, 49, 400–409.
- Gibbons, J. W., Scott, D. E., Ryan, T. J., Buhlmann, K. A., Tuberville, T. D., Metts, B. S., Greene, J. L., Mills, T., Leiden, Y., Poppy, S., & Winne, C. T. (2000). The global decline of reptiles, déjà vu amphibians. BioScience, 50, 653-666.
- Glorioso, B. M., Vaughn, A. J., & Waddle, J. H. (2010). The aquatic turtle assemblage inhabiting a highly altered landscape in southeast Missouri. *Journal of Fish and Wildlife Management*, 1, 161–168. https://doi.org/10.3996/072010-JFWM-020
- Gross, J., Elvinger, F., Hungerford, L. L., & Gehrt, S. D. (2012). Raccoon use of the urban matrix in the Baltimore metropolitan area, Maryland. *Urban Ecosystems*, 15, 667–682. https://doi.org/10.1007/s1125 2-011-0218-z
- Gulette, A. L. (2018). Habitat suitability of restored wetlands and an investigation of sampling bias for freshwater turtles in West Virginia. Thesis, West Virginia University, Morgantown, West Virginia, USA.

- Gulette, A. L., Anderson, J. T., & Brown, D. J. (2019). Influence of hoopnet trap diameter and size distribution of comparatively large and small freshwater turtles. Northeastern Naturalist, 26, 129–136.
- Hairston, N. G. (1986). Species packing in *Desmognathus* salamanders: Experimental demonstration of predation and competition. *American Naturalist*, 127, 266–291. https://doi.org/10.1086/284485
- Hak, J. C., & Comer, P. J. (2017). Modeling landscape condition for biodiversity assessment-Application in temperate North America. *Ecological Indicators*, 82, 206–216. https://doi.org/10.1016/j.ecoli nd.2017.06.049
- Halsey, L. G. (2019). The reign of the p-value is over: What alternative analyses could we employ to fill the power vacuum? *Biology Letters*, 15, 20190174.
- Hardin, G. (1960). The competitive exclusion principle. *Science*, 131, 1292–1297. https://doi.org/10.1126/science.131.3409.1292
- Héritier, L., Valdeón, A., Sadaoui, A., Gendre, T., Ficheux, S., Bouamer, S., Kechemir-Issad, N., Du Preez, L., Palacios, C., & Verneau, O. (2017). Introduction and invasion of the red-eared slider and its parasites in freshwater systems of southern Europe: Risk assessment for the European pond turtle in wild environments. *Biodiversity and Conservation*, 26, 1817–1843.
- Hohn, S. M. (2003). A survey of New York state pet stores to investigate trade in native herpetofauna. *Herpetological Review*, 34, 23–27.
- Holderegger, R., & Di Giulio, M. (2010). The genetic effects of roads: A review of empirical evidence. *Basic and Applied Ecology*, 11, 522–531. https://doi.org/10.1016/j.baae.2010.06.006
- Howell, H. J., Legere, R. H. Jr, Holland, D. S., & Seigel, R. A. (2019). Long-term turtle declines: Protected is a verb, not an outcome. *Copeia*, 107, 493–501. https://doi.org/10.1643/CH-19-177
- Iverson, J. B., & Lewis, E. L. (2018). How to measure a turtle. *Herpetological Review*, 49, 453–460.
- Janzen, F. J., Paukstis, G. L., & Brodie, E. D. III (1992). Observations on basking behavior of hatchling turtles in the wild. *Journal of Herpetology*, 26, 217-219. https://doi.org/10.2307/1564866
- Jenks, G. F. (1977). Optimal data classification for choropleth maps. Occasional paper No. 2. Department of Geography, University of Kansas, Lawrence, USA.
- Kakuda, A., Doi, H., Souma, R., Nagano, M., Minamoto, T., & Katano, I. (2019). Environmental DNA detection and quantification of invasive red-eared sliders, *Trachemys scripta elegans*, in ponds and the influence of water quality. *PeerJ*, 7, e8155.
- Kennish, M. J. (2002). Environmental threats and environmental future of estuaries. *Environmental Conservation*, 29, 78–107. https://doi. org/10.1017/S0376892902000061
- Khera, N., Das, A., Srivasatava, S., & Jain, S. (2010). Habitat-wise distribution of the House Sparrow (*Passer domesticus*) in Delhi, India. *Urban Ecosystems*, 13, 147–154. https://doi.org/10.1007/s1125 2-009-0109-8
- Lambert, M. R., McKenzie, J. M., Screen, R. M., Clause, A. G., Johnson, B. B., Mount, G. G., Shaffer, H. B., & Pauly, G. B. (2019). Experimental removal of introduced slider turtles offers new insight into competition with a native, threatened turtle. *PeerJ*, 7, e7444. https://doi.org/10.7717/peerj.7444
- Lefevre, K., & Brooks, R. J. (1995). Effects of sex and body size on basking behavior in a northern population of the painted turtle, *Chrysemys picta*. *Herpetologica*, *51*, 217–224.
- Leu, M., Hanser, S. E., & Knick, S. T. (2008). The human footprint in the west: A large-scale analysis of anthropogenic impacts. *Ecological Applications*, 18, 1119–1139. https://doi.org/10.1890/07-0480.1
- Leveau, L. M. (2018). Urbanization, environmental stabilization and temporal persistence of bird species: A view from Latin America. *PeerJ*, 6, e6056.
- Li, B., Zhang, W., Shu, X., Pei, E., Yuan, X., Sun, Y., Wang, T., & Wang, Z. (2016). The impacts of urbanization on the distribution and body condition of the rice-paddy frog (Fejervarya multistriata) and gold-striped

- pond frog (Pelophylax plancyi) in Shanghai, China. Asian Herpetological Research, 7, 200–209.
- Lindeman, P. V. (1999). Aggressive interactions during basking among four species of emydid turtles. *Journal of Herpetology*, 33, 214–219. https://doi.org/10.2307/1565717
- Longcore, T., & Rich, C. (2004). Ecological light pollution. Frontiers in Ecology and the Environment, 2, 191–198.
- Lovich, J. E., Ennen, J. R., Agha, M., & Gibbons, J. W. (2018). Where have the turtles gone, and why does it matter? *BioScience*, 68, 771–781.
- Lowe, S., Browne, M., Boudjelas, S., & De Poorter, M. (2000). 100 of the world's worst invasive alien species: A selection from the global invasive species database. Invasive Species Specialist Group, International Union for the Conservation of Nature. http://www.iucngisd.org/ gisd/pdf/100English.pdf
- Maceda-Veiga, A., Green, A. J., & de Sosta, A. (2014). Scaled body-mass index shows how habitat quality influences the condition of four fish taxa in north-eastern Spain and provides a novel indicator of ecosystem health. Freshwater Biology, 59, 1145–1160. https://doi. org/10.1111/fwb.12336
- Mali, I., Brown, D. J., Ferrato, J. R., & Forstner, M. R. J. (2014). Sampling freshwater turtle populations using hoop nets: Testing potential biases. Wildlife Society Bulletin, 28, 580–585. https://doi.org/10.1002/ wsb.427
- Mali, I., Dickerson, B. E., Brown, D. J., Dixon, J. R., & Forstner, M. R. J. (2013). Road density not a major driver of Red-Eared Slider (*Trachemys scripta elegans*) population demographics in the lower Rio Grande Valley of Texas. *Herpetological Conservation and Biology*, 8, 131–140.
- McCormick, P. V., & Laing, J. A. (2003). Effects of increased phosphorus loading on dissolved oxygen in a subtropical wetland, the Florida Everglades. *Wetlands Ecology and Management*, 11, 199–216.
- McGoff, E., Solimini, A. G., Pusch, M. T., Jurca, T., & Sandin, L. (2013). Does lake habitat alteration and land-use pressure homogenize European littoral macroinvertebrate communities? *Journal of Applied Ecology*, 50, 1010–1018. https://doi.org/10.1111/1365-2664.12106
- McGowan, K. J. (2001). Demographic and behavioral comparisons of suburban and rural American crows. In J. M. Marzluff, R. Bowman, & R. Donnelly (Eds.), Avian ecology and conservation in an urbanizing world (pp. 365–381). Springer.
- McKinney, M. L., & Lockwood, J. L. (1999). Biotic homogenization: A few winners replacing many losers in the next mass extinction. *Trends in Ecology & Evolution*, 14, 450–453. https://doi.org/10.1016/S0169-5347(99)01679-1
- Menge, B. A. (1976). Organization of the New England rocky tidal community: Role of predation, competition, and environmental heterogeneity. *Ecological Monographs*, 46, 355–393.
- Miles, L. S., Breitbart, S. T., Wagner, H. H., & Johnson, M. T. J. (2019). Urbanization shapes the ecology and evolution of plant-arthropod herbivore interactions. Frontiers in Ecology and Evolution, 7, 310. https://doi.org/10.3389/fevo.2019.00310
- Miraldo, A., Li, S., Borregaard, M. K., Flórez-Rodríguez, A., Gopalakrishnan, S., Rizvanovic, M., Wang, Z., Rahbek, C., Marske, K. A., & Nogués-Bravo, D. (2016). An Anthropocene map of genetic diversity. *Science*, 353, 1532–1535. https://doi.org/10.1126/science.aaf4381
- Morreale, S. J., & Gibbons, J. W. (1986). Habitat suitability index models: Slider turtle. U.S. Fish and Wildlife Service Biological Report 82, Washington, D.C., USA.
- Mullin, D. I., White, R. C., Lentini, A. M., Brooks, R. J., Bériault, K. R., & Litzgus, J. D. (2020). Predation and disease limit population recovery following 15 years of headstarting an endangered freshwater turtle. Biological Conservation, 245, 108496.
- Murray, L. G., & Seed, R. (2010). Determining whether catch per unit effort is a suitable proxy for relative crab abundance. Marine Ecology Progress Series, 401, 173–182. https://doi.org/10.3354/meps08415

- Naselli-Flores, L., & Barone, R. (2000). Phytoplankton dynamics and structure: A comparative analysis in natural and man-made water bodies of different trophic states. *Hydrobiologia*, 438, 65–74.
- Oh, H., Park, S., Adhikari, P., Kim, Y., Kim, T., & Han, S. (2017). Distribution and status of the alien invasive red-eared slider (*Trachemys scripta elegans*) in Jeju Island, South Korea. *Korean Journal of Environmental Biology*, 35, 57-63. https://doi.org/10.11626/KJEB.2017.35.1.057
- Ordonez, A., Martinuzzi, S., Radeloff, V. C., & Williams, J. W. (2014). Combined speeds of climate and land-use change of the conterminous US until 2050. *Nature Climate Change*, 4, 811–816. https://doi.org/10.1038/nclimate2337
- Ordóñez, C., & Duinker, P. N. (2012). Ecological integrity in urban forests. *Urban Ecosystems*, 15, 863–877. https://doi.org/10.1007/s1125 2-012-0235-6
- Oxenrider, K. J., Heres, B. M., Mota, J. L., & Brown, D. J. (2019). Influence of bait type on capture success of *Clemmys guttata* and *Chrysemys picta* using small hoop nets in shallow wetlands. *Herpetological Review*, 50, 490–492.
- Parrish, J. D., Braun, D. P., & Unnasch, R. S. (2003). Are we conserving what we say we are? Measuring ecological integrity within protected areas. *BioScience*, 53, 851–860.
- Peltzer, P. M., Lajmanovich, R. C., Attademi, A. M., & Beltzer, A. H. (2006). Diversity of anurans across agricultural ponds in Argentina. Biodiversity and Conservation, 15, 3499–3513. https://doi.org/10.1007/s10531-004-2940-9
- Petrozzi, F., Ajong, S. N., Pacini, N., Dendi, D., Bi, S. G., Fa, J. E., & Luselli, L. (2021). Spatial niche expansion at multiple habitat scales of a tropical freshwater turtle in the absence of a potential competitor. *Diversity*, 13, 55. https://doi.org/10.3390/d13020055
- Polo-Cavia, N., López, P., & Martín, J. (2011). Aggressive interactions during feeding between native and invasive freshwater turtles. *Biological Invasions*, 13, 1387–1396. https://doi.org/10.1007/s10530-010-9897-2
- Prange, S., Gehrt, S. D., & Wiggers, E. P. (2003). Demographic factors contributing to high raccoon densities in urban landscapes. *Journal of Wildlife Management*, 67, 324–333. https://doi. org/10.2307/3802774
- Pulliam, H. R. (2000). On the relationship between niche and distribution. *Ecology Letters*, 3, 349–361. https://doi.org/10.1046/j.1461-0248.2000.00143.x
- Riley, S. P. D., Hadidian, J., & Manski, D. A. (1998). Population density, survival, and rabies in raccoons in an urban national park. *Canadian Journal of Zoology*, 76, 1153–1164. https://doi.org/10.1139/z98-042
- Rodewald, A. D., Kearns, L. J., & Shustack, D. P. (2011). Anthropogenic resource subsidies decouple predator-prey relationships. *Ecological Applications*, 21, 936–943. https://doi.org/10.1890/10-0863.1
- Roseberry, J. L., & Woolf, A. (1998). Habitat-population density relationships for white-tailed deer in Illinois. Wildlife Society Bulletin, 26, 252–258.
- Rosenfield, R. N., Bielefeldt, J., Affeldt, J. L., & Beckman, D. J. (1995).
 Nesting Diversity, nest area reoccupancy, and monitoring implications for Cooper's hawks in Wisconsin. *Journal of Raptor Research*, 29 1-4
- Ryan, T. S., Conner, C. A., Douthitt, B. A., Sterrett, S. C., & Salsbury, C. M. (2008). Movement and habitat use of two aquatic turtles (*Graptemys geographica* and *Trachemys scripta*) in an urban landscape. *Urban Ecosystems*, 11, 213–225. https://doi.org/10.1007/s1125 2-008-0049-8
- Sala, O. E., Chapin, F. S. III, Armesto, J. J., Berlow, E., Bloomfield, J., Dirzo, R., Huber-Sanwald, E., Huenneke, L. F., Jackson, R. B., Kinzig, A., Leemans, R., Lodge, D. M., Mooney, H. A., Oesterheld, M., Poff, N. L. R., Sykes, M. T., Walker, B. H., Walker, M., & Wall, D. H. (2000). Global biodiversity scenarios for the year 2100. Science, 287, 1770–1774.
- Sasaki, K., Lesbarrères, D., Beaulieu, C. T., Watson, G., & Litzgus, J. (2016). Effects of a mining-altered environment on individual

- fitness of amphibians and reptiles. *Ecosphere*, 7, e01360. https://doi.org/10.1002/ecs2.1360
- Schulte-Hostedde, A. I., Zinner, B., Millar, J. S., & Hickling, G. J. (2005). Restitution of mass-size residuals: Validating body condition indices. *Ecology*, 86, 155–163. https://doi.org/10.1890/04-0232
- Segre, H., DeMalach, N., Henkin, Z., & Kadmon, R. (2016). Quantifying competitive exclusion and competitive release in ecological communities: A conceptual framework and case study. PLoS One, 11, e0160798.
- Slabbekoorn, H., & Ripmeester, E. A. P. (2008). Birdsong and anthropogenic noise: Implications and applications for conservation. *Molecular Ecology*, 17, 72–83. https://doi.org/10.1111/j.1365-294X.2007.03487.x
- Smith, V. H., & Schindler, D. W. (2009). Eutrophication science: Where do we go from here? Trends in Ecology and Evolution, 24, 201–207. https://doi.org/10.1016/j.tree.2008.11.009
- Sofaer, H. R., Flather, C. H., Jarnevich, C. S., Davis, K. P., & Pejchar, L. (2020). Human-associated species dominate passerine communities across the United States. *Global Ecology and Biogeography*, 29, 885–895. https://doi.org/10.1111/geb.13071
- Spencer, R.-J., & Thompson, M. B. (2005). Experimental analysis of the impact of foxes on freshwater turtle populations. *Conservation Biology*, 19, 845–854. https://doi.org/10.1111/j.1523-1739.2005.00487.x
- Stone, P. A., Powers, S. M., & Babb, M. E. (2005). Freshwater turtle assemblages in central Oklahoma farm ponds. Southwestern Naturalist, 50, 166–171.
- Telecky, T. M. (2001). United States import and export of live turtles and tortoises. *Turtle and Tortoise Newsletter*, 4, 8–13.
- Traweger, D., Travnitzky, R., Moser, C., Walzer, C., & Bernatzky, G. (2006). Habitat preferences and distribution of the brown rat (*Rattus norvegicus* Berk.) in the city of Salzburg (Austria): Implications for an urban rat management. *Journal of Pest Science*, 79, 113–125.
- Tucker, J. K., & Lamer, J. T. (2008). Homing in red-eared slider (*Trachemys scripta elegans*) in Illinois. Chelonian Conservation and Biology, 7, 145–149.

- Watson, M. B., & Pauley, T. K. (2006). Spatial distribution of turtles along the Great Kanawha River, West Virginia. *Proceedings of the West* Virginia Academy of Science, 78, 14–25.
- West, E. H., Henry, W. R., Goldenberg, W., & Peery, M. Z. (2016). Influence of food subsidies on the foraging ecology of a synanthropic species in protected areas. *Ecosphere*, 7, e01532. https://doi.org/10.1002/ecs2.1532
- Wilberg, M. J., Livings, M. E., Barkman, J. S., Morris, B. T., & Robinson, J. M. (2011). Overfishing, disease, habitat loss, and potential extirpation of oysters in upper Chesapeake Bay. *Marine Ecology Progress Series*, 436, 131–144. https://doi.org/10.3354/meps09161
- Zuur, A. F., Ieno, E. N., & Elphick, C. S. (2010). A protocol for data exploration to avoid common statistical problems. *Methods in Ecology and Evolution*, 1, 3–14. https://doi.org/10.1111/j.2041-210X.2009.00001.x
- Zuur, A. F., Ieno, E. N., Walker, N. J., Saveliev, A. A., & Smith, G. M. (2009). Mixed effects models and extensions in ecology with R. Springer Science + Business Media.
- Zwick, P. (1992). Stream habitat fragmentation a threat to biodiversity. *Biodiversity and Conservation*, 1, 80–97. https://doi.org/10.1007/BF00731036

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

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